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PREPARATIVE UV-LASER PHOTOCHEMISTRY OF THE AZOALKANE SPIRO(2,3-DIAZABICYCLO-[2,2,1]HEPT-2-ENE-7',1-CYCLOPROPANE):

TRAPPING OF THE 1,4-Diradical 2-(3-Cyclopentenyl)ethyl by Molecular Oxygen

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<u>SUMMARY</u>: Photo-extrusion of nitrogen from the azoalkane $\frac{1}{2}$ in the presence of molecular oxygen gave besides the hydrocarbons $\frac{3}{2}$ and $\frac{5}{2}$, the endoperoxide $\frac{10}{2}$ and hydroperoxide $\frac{11}{2}$, the former via trapping of the 1,4-diradical $\frac{4}{2}$ by triplet oxygen, the latter by ene-reaction of hydrocarbon $\frac{5}{2}$ with singlet oxygen.

The photolysis of the azoalkane $\underline{1}$ affords the two hydrocarbons spiro(bicyclo[2.1.0]pentane-5',1-cyclopropane) ($\underline{3}$) and bicyclo[3.2.0]hept-1-ene ($\underline{5}$), postulated to be derived from the 1,3diradical spiro[2.4]hepta-4,7-diyl ($\underline{2}$) and the 1,4-diradical 2-(3-cyclopentenyl)ethyl ($\underline{4}$), respectively, as illustrated in Eq.1.¹ Direct photolysis gave predominantly the cyclization product $\underline{3}$, while benzophenone-sensitized photolysis gave mainly the rearranged product $\underline{5}$. To



rationalize this spin state selectivity, it was argued¹ that in its singlet state the 1,3diradical $\frac{2}{2}$ preferred cyclization into $\frac{3}{2}$, but in its triplet state ring-opening to the 1,4diradical $\frac{4}{2}$ predominated, followed by cyclization into $\frac{5}{2}$. Stereolabeling experiments of the cyclopropane ring in azoalkane $\frac{1}{2}$ showed that the intervention of the 1,4-diradical $\frac{4}{2}$ was plausible.

The recent success of trapping 1,3-diradicals $\underline{12} - \underline{14}^{2,3}$ and 1,4-diradicals $\underline{15} - \underline{12}^4$ by molecular oxygen to form of the corresponding peroxides encouraged us to apply this useful



diradical probe in the photolysis of azoalkane $\underline{1}$. The spirocyclopropane moiety in diradical $\underline{2}$ was particularly interesting and significant in that the built-in cyclopropylcarbinyl "free radical clock"⁵ provided us with a means of estimating lifetime limits of the transient species of the photolysis of azoalkane $\underline{1}$. We anticipated that the triplet 1,3-diradical $\underline{2}$ would ring-open faster to the triplet 1,4-diradical $\underline{4}$ than being trapped with triplet oxygen to form endoperoxide $\underline{7}$, but that oxygen trapping of the 1,4-diradical $\underline{4}$ would give the 1,2-dioxane $\underline{10}$ faster than cyclization into the hydrocarbon $\underline{5}$. The need of monochromicity at high intensities for the triplet sensitization and the expected short lifetimes of the diradical intermediates, especially the spiro[2.4]hepta-4,7-diyl ($\underline{2}$), dictated making use of laser photochemistry at elevated oxygen pressure $\underline{4a}$ to optimize diradical trapping.

Irradiation of a CFCl₃ solution of azoalkane $\underline{1}$ (0.1 \underline{M}) and benzophenone (0.13 \underline{M}), contained in a Griffin-Worden tube under a 10 atm pressure of oxygen gas, was carried out at -20°C with the 363.7-nm line (ca. 1 Watt intensity) of a Coherent Supergraphite Model CR-18 argon ion laser for 3 h. TLC(silica gel, CH₂Cl₂) monitoring revealed the presence of two peroxides, spotted by means of the ferrous thiocyanate spray.⁶ Silica gel chromatography at -20°C and eluting with CH₂Cl₂ gave 8% of a mixture of hydrocarbons $\underline{3}$ and $\underline{5}$, 30% unreacted azoalkane $\underline{1}$, the 1,2-dioxane $\underline{10}^7$ and the hydroperoxide $\underline{11}^8$, in 21% and 32% yields, respectively. Structural assignment of these peroxides rested on positive peroxide test, elemental composition and spectra data, particularly the characteristic ¹³C- and ¹H-NMR spectra. The latter were exceedingly complex, but with the help of decoupling and computer simulation the observed spectra were consistent with the proposed structures. In addition, the hydroperoxide $\underline{11}$ could be independently prepared in 74% yield by tetraphenylporphine-sensitized photooxygenation of hydrocarbon $\underline{5}$ in CH₂Cl₂ at -20°C.

Control experiments revealed that not even traces of the endoperoxide $\underline{\underline{7}}$ could be detected in the laser photolysis mixture of azoalkane $\underline{1}$. For this purpose authentic endoperoxide $\underline{\underline{7}}$ was prepared by photooxygenation and dimide reduction.⁹ Under the benzophenone-sensitized conditions the hydrocarbon $\underline{5}$ was converted to the hydroperoxide $\underline{11}$, but in the dark the hydrocarbon $\underline{5}$ was stable towards triplet oxygen. Thus, the hydroperoxide $\underline{11}$ was the product of the ene-reaction of hydrocarbon $\underline{5}$ with in situ singlet oxygen that was generated via quenching of benzophenone triplets by oxygen.¹⁰

The bona fide oxygen trapping product in the laser photolysis of azoalkane $\underline{1}$ was the 1,2dioxane $\underline{10}$. The hydroperoxide $\underline{11}$ was a secondary product, derived from singlet oxygenation of of hydrocarbon $\underline{5}$. Since in the laser photolysis ca. 32% hydroperoxide $\underline{11}$ and ca. 8% hydrocarbons $\underline{3}$ and $\underline{5}$ were formed, the effective yield of the latter is ca. 40%. Thus, the 1,4-diradical $\underline{4}$ predominantly cyclizes to hydrocarbon $\underline{5}$, but oxygen trapping is competitive since substantial amounts (ca. 21%) of 1,2-dioxane $\underline{10}$ were isolated. Since no endoperoxide $\underline{7}$ could be detected, the 1,3-diradical $\underline{2}$ must be significantly shorter lived (about 10- to 100-fold) than the 1,4-diradical $\underline{4}$. Trapping of diradicals by oxygen is essentially diffusion controlled, within the confines of spin statistics as evidenced by the measured bimolecular rate constant k= 2.4 x 10⁹ m⁻¹sec⁻¹.¹¹ Thus, under the trapping conditions employed here, the lifetime (τ) of the 1,4-diradical $\underline{4}$ must be at least 1 nsec and consequently that of the 1,3-diradical $\underline{2}$ less than 0.1 nsec, possibly as short as 0.01 nsec.

Of the two possible trapping routes $\frac{4}{2} + \frac{8}{2} + \frac{10}{2}$ and $\frac{4}{2} + \frac{9}{2} + \frac{10}{2}$ (Eq.1) clearly the latter is preferred in view of the allylic stabilization in the 1,6-diradical $\frac{9}{2}$, which is absent in the 1,6-diradical $\frac{8}{2}$. However, the latter could in principle arise via the path $\frac{2}{2} + \frac{6}{2} + \frac{8}{2}$. In view of the fact that the rate constant of the cyclopropylcarbinyl rearrangement is only 2.0 x 10⁸ sec⁻¹, ⁵ the ring-opening of the 1,5-diradical $\frac{6}{2}$ is much too slow compared to oxygen trapping and cyclization. We should have detected endoperoxide $\frac{7}{2}$ under the conditions employed here. Thus, the $\frac{2}{2} + \frac{6}{2} + \frac{8}{2} + \frac{10}{2}$ route cannot be important because the 1,3-diradical $\frac{2}{2}$ is too short-lived on account of its facile ring-opening to the 1,4-diradical $\frac{4}{2}$. The gain in allylic stabilization and the release in cyclopropane strain must be responsible for the very fast (ca. 10¹¹ sec⁻¹) ring-opening $\frac{2}{2} + \frac{4}{2}$.

The present investigation demonstrates that diradical trapping by molecular oxygen under laser photolysis conditions provides a powerful mechanistic tool for characterizing such elusive intermediates and estimating their lifetimes. Particularly advantageous for this purpose is the monochromatic and intense light output of the argon ion laser¹², optimal requisites for generating and trapping diradical intermediates.

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- 7. Endoperoxide $\underline{10}$, colorless liquid, b.p. $100^{\circ}C/14$ Torr (Kugelrohr).-IR (CCl₄): 3050, 2950, 2920, 2870, 1460, 1440, 1435, 1355, 1040, 1010 cm⁻¹.-¹H-NMR (CDCl₃), 400 MHz: $\delta = 1.49 - 1.59$ (m, J_{1,9} = 6.4, J_{8,9} = 10.1, J_{8',9'} = 6.7, J_{9,9'} = 13.4 Hz; 1H, 9'-H), 2.21 - 2.30 (m, J_{1,9} = 7.9, J_{8,9} = 1.8, J_{8',9} = 8.1 Hz; 1H, 9-H), 2.30 - 2.46 (m, J_{1,8} = 2, J_{1,8'} = 1.8, J_{5,8} = 2.5, J_{5,8'} \sim 3, J_{7,8} = 2.3, J_{7,8'} = 2.3, J_{8,8'} = 13.5 Hz; 2H, 8-H and 8'-H), 2.48 - 2.53 (m, J_{1,5'} = 0.6, J_{4,5'} = 1.0, J_{4',5'} = 3.0, J_{5,5'} = 13.5, J_{7,5'} = 0.4 Hz; 1H, 5'-H), 2.60 - 2.70 (m, J_{4,5} = 6.1, J_{4',5} = 12.2, J_{7,5} = 1.8 Hz; 1H, 5-H), 4.07 - 4.21 (m, J_{4,4'} = 12.2 Hz; 2H, 4-H and 4'-H), 5.12 - 5.17 (m; 1H, 1-H), 5.59 - 5.63 (m, J_{1,7} = 0.8 Hz; 1H, 7-H).-¹³C-NMR (CDCl₃), 100.6 MHz: $\delta = 27.3$ (t), 27.6 (t), 29.2 (t), 72.9 (t), 88.4 (d), 125.2 (d), 136.9 (s).-MS (70 eV): m/e = 126 (6%, M⁺), 108 (20%), 97 (46%), 80 (20%), 79 (100%), 77 (66%), 41 (44%).-C₇H₁₀O₂ (126.12): Calcd. C 66.64 H 7.99 Found C 66.69 H 7.96
- 8. Hydroperoxide 11, colorless liquid, b.p. 20°/0.1 Torr (Kugelrohr).-

IR (CC1₄): 3590, 3110, 3040, 2990, 2960, 2890, 1445, 1370, 1330, 1250 cm⁻¹.-¹H-NMR (CDC1₃), 400 MHz: $\delta = 1.09 - 1.19$ (m, J_{6,6}⁻ = 12.1, J₆⁻, 7⁻ ⁰ 10, J₆⁻, 7 = 9.7, J_{5,6}⁻ = 5.4, J_{4,6}⁻ = 0.4, J_{2,6}⁻ = 0.4 Hz; 1H, 6⁻-H), 2.00 - 2.18 (m, J_{7,7}⁻ ⁰ 12, J_{6,7}⁻ ⁰ 9, J_{6,7}⁻ ⁰ 9.5 Hz; 3H, 4⁻-H, 6-H and 7⁺-H), 2.35 - 2.45 (m, J_{4,7} = 1, J_{5,7} = 0.4 Hz; 1H, 7-H), 2.61 -2.71 (m, J_{4,4}⁻ = 17.0, J_{4,5} = 6.9, J_{3,4} = 1.8, J_{2,4} = 2.7 Hz; 1H, 4-H), 2.79 - 2.89 (m, J₄⁺, 5 = 0.6, J_{5,6}⁻ ⁰ 9, J_{3,5} = 0.9, J_{2,5} = 0.7 Hz; 1H, 5-H), 5.71 - 5.75 (m, J_{2,3} = 5.5, J_{2,4}⁻ = 1.8 Hz; 1H, 2-H), 6.11 - 6.15 (m, J_{3,4}⁻ = 2.5 Hz; 1H, 3-H), 7.87 (br.s, 1H, -00H).-¹³C-NMR (CDC1₃), 100.6 MHz: $\delta = 20.1$ (t), 30.4 (t), 39.6 (t), 39.5 (d), 96.5 (s), 130.0 (d), 137.9 (d).-MS (70 eV): m/e = 126 (1%, M⁺), 110 (22%), 82 (95%), 79 (55%), 68 (45%), 54 (47%), 39 (100%).-C₇H₁₀O₂ (126.16): Calcd. C 66.64 H 7.99 Found C 66.50 H 8.18

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